

KUWAITIMYCIN, EFFECT ON SYNTHESIS OF LIPIDS IN
BACILLUS SUBTILIS CELLS

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The effects exerted by kuwaitimycin on synthesis of lipids as well as some metabolic activities of *Bacillus subtilis* were studied. The antibiotic not only arrested the incorporation of ^{14}C -acetate into the microbial lipids but also altered the fatty acids pattern, contents of i-C 15, a-C 15, i-C 17 and a-C 17 were markedly reduced, concomitant with an increase in the contents of i-C 14 and n-C 14. Moreover, the rates of synthesis of phospholipids were decreased by the drug, especially that of phosphatidyl ethanolamine.

Kuwaitimycin is a peptolide antibiotic containing several amino acids and isohexadeca- 3,6 dienoic acid as the fatty acid moiety¹⁾ of the molecule.

The effect of the antibiotic on transport of alkali earth metal ions through mitochondrial membrane was demonstrated by SHIMI²⁾.

During the present work the probable effects exerted by kuwaitimycin on synthesis of lipids and some other metabolic activities of *B. subtilis* were explored in order to locate its primary site of action on the microbial test organism.

Materials and Methods

Kuwaitimycin was prepared and purified as described by SHIMI *et al*³⁾.

Bacillus subtilis was grown in nutrient broth having the following composition (g/100 ml): peptone, 0.5; NaCl, 0.5, meat extract, 0.15 and yeast extract 0.15. Cultures were shaken at 220 rpm at 35°C and the bacterial cells were then harvested at $15,000 \times g$ during the middle logarithmic phase of growth. The harvest was diluted with sterile medium to produce a bacterial suspension of O.D. 0.4 at 660 nm. This preparation was used throughout the following experiments.

The effect of kuwaitimycin on the biosynthesis of macromolecules was carried out as described before³⁾.

Effect of Kuwaitimycin on Lipid Synthesis of *B. subtilis*

Lipid synthesis in medium containing U- ^{14}C -acetate (58 mCi/m mol) with and without kuwaitimycin was investigated. Details of experimental conditions were as follows: 5 ml samples of the cultures were withdrawn at appropriate incubation periods and the bacterial cells were harvested by centrifugation. The lipid content of the harvested cells was extracted as described by BLIGH & DYER⁴⁾. A portion of each lipid extract was used for radioactivity measurements using a liquid scintillation spectrometer Nuclear Chicago Mark II in a toluene-based PPO-POPOP system. The rest of the lipid extract was used for identification of phospholipids by TLC using chloroform - methanol - acetic acid (65: 25: 10, v/v) as developing solvent. Location of spots on developed chromatograms was carried out by comparison with authentic samples of phospholipids. The radiochemicals were purchased from the Radiochemical Centre, Amersham, Bucks, U.K.

Effect of Kuwaitimycin on Fatty Acid Synthesis in *B. subtilis* Cells

Equal portions of the bacterial preparation were separately diluted with equal volumes of the fresh warm sterile medium containing different concentrations of kuwaitimycin and then incubated for different periods at 37°C. The bacterial cells were harvested by centrifugation and their lipid con-

tents were extracted⁴⁾, methylated⁵⁾, and analysed by GLC using a 20% PEGA column. Results and experimental details are given in Table 1 and the legend.

Results and Discussion

The effect of kuwaitimycin on *B. subtilis* growth is given in Fig. 1. Kuwaitimycin did not exert significant effects on the synthesis of labeled macromolecules at early incubation periods.

Kuwaitimycin partially arrested the utilization of ¹⁴C-acetate by the organism (Fig. 2). The rate

Fig. 1. Effect of kuwaitimycin on the growth of *B. subtilis*

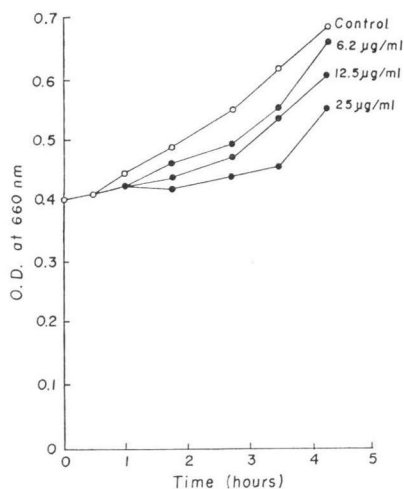


Fig. 2. Effect of kuwaitimycin on the incorporation of U-¹⁴C-acetate into *B. subtilis* lipids

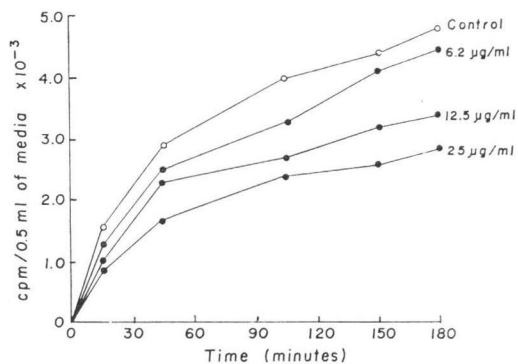
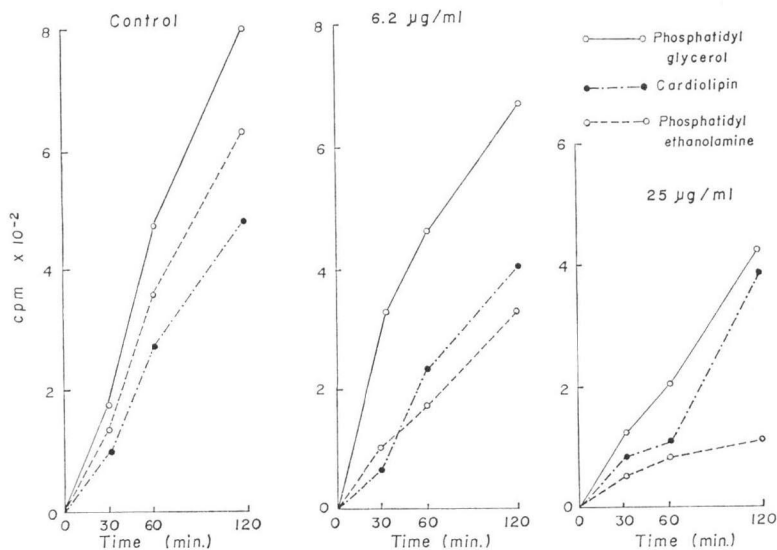


Fig. 3. Effect of kuwaitimycin on the phospholipid synthesis in *B. subtilis* cells

The identified phospholipid spots were scraped off from the TLC plates and their radioactivity was counted in polyethylene vials using the scintillation cocktail given in the text containing 4% (w/v) Cab-o-Sil.



of synthesis of phosphatidyl ethanolamine was markedly reduced at the MIC level of the antibiotic when compared with the control data, whereas the synthesis of cardiolipin and phosphatidyl glycerol were less markedly affected (Fig. 3). Similar results were also reported for phenethyl alcohol⁶⁾, polyketoacidomycin⁷⁾ and polymyxins⁸⁾. The decrease in the rates of synthesis of phospholipids became more pronounced the higher the concentration of the antibiotic applied (Fig. 3). Moreover, the relative contents of the fatty acids of C-15 and C-17 were reduced while the contents of i-C 14 and n-C 14 were increased (Table 1). RINGRASE & HIGGINS⁹⁾ reported similar findings in fatty acids pattern of *E. coli* treated cells.

Release of Na⁺ and K⁺ ions from inside the bacterial cells (Table 2) was augmented by kuwaiti-

Table 1. Effect of kuwaitimycin on the fatty acid pattern of *B. subtilis* cells

Fatty acids*	Concentration of fatty acids ($\mu\text{g}/\text{mg}$ cell dry weight)											
	15 minutes			40 minutes			60 minutes			120 minutes		
	Concentration of kuwaitimycin ($\mu\text{g}/\text{ml}$)											
	0.00	6.2	12.5	0.00	6.2	12.5	0.00	6.2	12.5	0.00	6.2	12.5
i-14	0.30	0.3	0.55	0.33	0.35	0.58	0.4	0.32	0.6	0.59	1.04	0.71
n-14	0.10	0.11	0.32	0.13	0.15	0.50	0.16	0.12	0.5	0.35	1.04	0.44
i-15	9.11	8.53	8.01	9.00	8.45	7.99	9.88	8.61	8.23	10.9	8.85	8.42
a-15	8.83	8.32	8.10	8.98	8.13	7.50	9.36	8.45	8.4	10.01	8.36	8.33
i-16	0.87	0.62	0.75	1.21	1.36	0.93	1.44	1.97	1.28	1.51	2.04	1.42
n-16	2.91	3.15	2.86	3.02	3.23	3.30	3.12	3.15	3.90	3.63	3.10	3.86
i-17	4.01	3.55	3.13	4.40	3.50	3.13	4.48	3.63	3.40	5.01	3.84	3.77
a-17	2.00	1.52	1.31	2.40	1.56	1.38	2.40	2.00	1.82	2.80	2.01	1.83

The fatty acid methyl esters are subjected to gas-liquid chromatography on 20% PEGA column at 185°C using N₂ as carrier gas (40 ml/minute). Analysis was carried out on PYE UNICAM GCV.

* n-14 & n-16=normal fatty acids of 14 & 16 carbon atoms respectively.

i-14, i-15, i-16 & i-17=iso fatty acids of 14, 15, 16 & 17 carbon atoms respectively.

a-15 & a-17=anteiso fatty acids of 15 & 17 carbon atoms respectively.

Table 2. Effect of kuwaitimycin on the intracellular concentration of Na and K ions

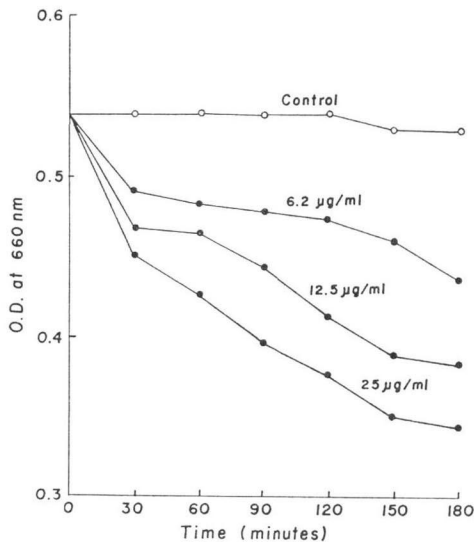
	Concentration of kuwaitimycin ($\mu\text{g}/\text{ml}$)					
	0.00		6.2		12.5	
	Cations concentration* ($\mu\text{g}/\text{mg}$ cell dry weight)					
	Na ⁺	K ⁺	Na ⁺	K ⁺	Na ⁺	K ⁺
15 min.	20	8	18	5	11	5
40 min.	83	37	58	23	50	18
60 min.	110	49	70	35	43	29
120 min.	115	43	70	30	39	30
240 min.	90	33	85	39	40	32

* Concentration of cations in the medium $\mu\text{g}/\text{ml}$: Na⁺ 210, K⁺ 70

B. subtilis cells were grown under the experimental conditions given in the text. Five ml samples of treated and non-treated control cultures were withdrawn at the preindicated incubation periods, washed with bidistilled water and the concentration of Na⁺ and K⁺ ions were determined using the Perkin Elmer atomic absorption spectrophotometer type 403 according to manufacturer's instructions (March 1971).

Fig. 4. Effect of kuwaitimycin on *B. subtilis* protoplasts

Washed cells of *B. subtilis* were suspended in a phosphate buffer (pH 7.2) containing 0.5 M sucrose and 100 $\mu\text{g}/\text{ml}$ of lysozyme. The suspension was incubated for 90 minutes at 30°C. The formation of protoplasts was confirmed microscopically. The effect of kuwaitimycin was determined as follows: 2.7 ml of a suspension of *B. subtilis* protoplasts (ca. 2×10^8 cell/ml), and 0.3 ml of solution containing the antibiotic were added to give a final volume of 3 ml. The mixture was incubated at 37°C and the turbidity at 660 nm was measured at the preindicated incubation periods.



mycin which can be attributed to changes in lipid contents of the bacterial cell membrane and consequent alteration of its permeability. Similar conclusions were reported in cases of bacitracin¹⁰⁾ and polymyxins¹¹⁾.

Kuwaitimycin caused bursting of *B. subtilis* protoplasts in hypertonic solutions probably due to changes in conformation and rigidity of cell membrane that can be attributed to variation in lipid composition (Fig. 4).

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